



## ANTIOXIDANT PROPERTIES AND THEIR DIVERSITY IN MUSHROOM MYCOFLORA

Dr. Vinay Kumar Singh

Associate professor, Mycology and Plant Pathology Laboratory,  
Department of Botany, K. S. Saket P.G. College Ayodhya (UP) India,  
Dr. Ram Manohar Lohia Avadh University, Ayodhya, (UP) India.

### ABSTRACT:

Mushroom is the well-known and famous Fungal group which has different economic value. It may be edible or inedible, cultivable, or wild, and medicinal or poisonous type. Edible mushroom has specific dietary components which increase the viability of cells by inhibiting free radicles. The process of inhibiting free radicle known Antioxidant properties which is highly determined in mushroom species. The specific antioxidant component of mushroom is phenolic compounds and total antioxidant activity of mushroom reported by the total phenolic content. Which binds with the free radicles and minimize apoptosis, cell death and finally ageing of cells. Antioxidant also reported to as synthetic way but it may have side effects. In natural mean of antioxidant, mushroom is common and known to use with specific test, flavor and aroma. Mushroom may have a lifesaving property as antioxidant property which also increase the durability of life and lifespan of human. The present article exclusively deals with the mushroom species and their antioxidant properties and also their useful roles.



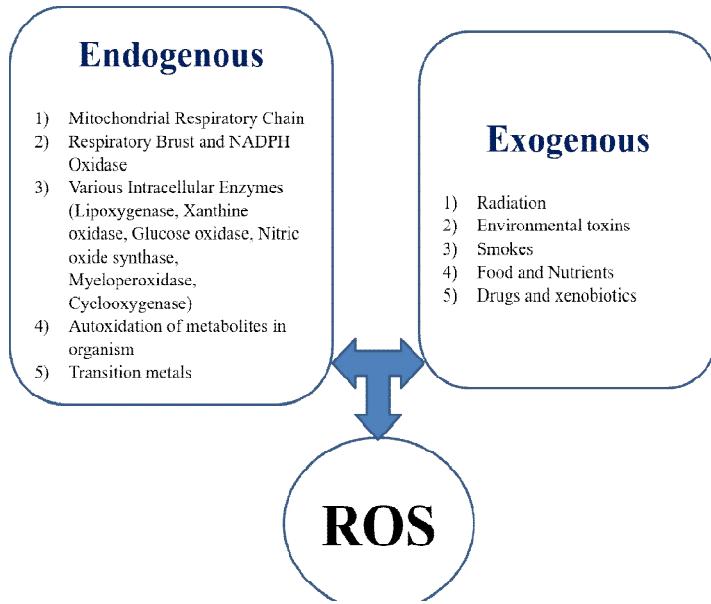
**KEYWORDS:** Mushroom, antioxidant, antioxidant activities, antioxidant system, free radicles, reactive oxygen species.

### INTRODUCTION:

Mushroom are distributed world widely and consume as dietary component from ancient time (Singh and Singh, 2022). Mushroom has its own specific test, flavor and aroma with edibility properties (Keles et al., 2011).Mushroom may have edible, medicinal and poisonous type. In the chemical properties of mushroom has 90% water and 10% of their matters. Edible mushroom has attractive nutritional value which can be compared with milk, egg, meat etc. Chemically mushroom contains several types of vitamins and essential amino acids, other than proteins, fats, carotenoids, organic acids etc. Energetic value of mushroom defined between 250 to 350 cal/kg. (Sanchez, 2017).Mushroom is a rich source of phenolic compounds, carotenoids, tocopherol and ascorbic acid. Mushroom has unlimited biological active agents as a source of useful therapeutical and about 700 species of mushrooms reported to have pharmacological activities (Ajith and Janardhanan, 2007).It is accumulating a variety of compounds including steroids, polyketides, terpenes and phenolic compounds as secondary metabolites. These compounds have antioxidant properties. Among all the antioxidant components, phenolic compounds have large array of biological actions which include free radical scavenging and inhibition of oxidation(Keles et al., 2011; Khatun et al., 2015).

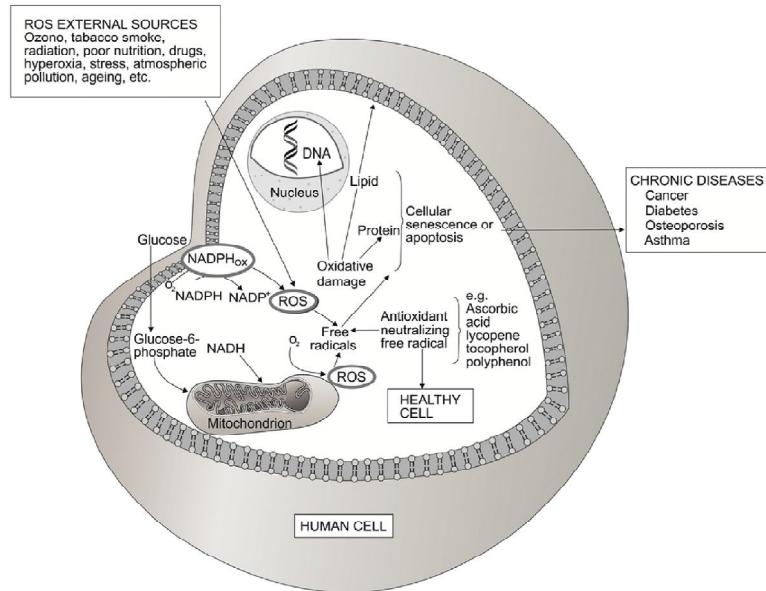
### Antioxidant System

It is estimated that molecular oxygen evolved about 2.45 billion years ago and introduced as O<sub>2</sub> in environment. These O<sub>2</sub> systematically evolving from photosynthetic organisms and reactive oxygen species (ROS) in aerobic life on earth. ROS may have two types with different components including indigenous and exogenous (Kosanic et al., 2013; Kump, 2008) (Fig. 1). The O<sub>2</sub> molecules has two free or impaired electrons known free radical which can damage the cells of all organisms. A free radical is a chemical compound that contains one or more unpaired electrons in atomic or molecular orbitals (Alliwell, 1994). Free radical is specified as reactive molecules derived from molecular oxygen. In animals specified into human, energy production and immune function performed by the process of oxidation. In normal physiological condition of body, low level of ROS produced by oxidation which maintain normal cell function and antioxidant defense system. Whereas the ROS level exceeds (in high concentration) from normal function, it expresses harmful action by damaging nucleic acids, protein oxidation and peroxidation of lipid (Sanchez, 2017).



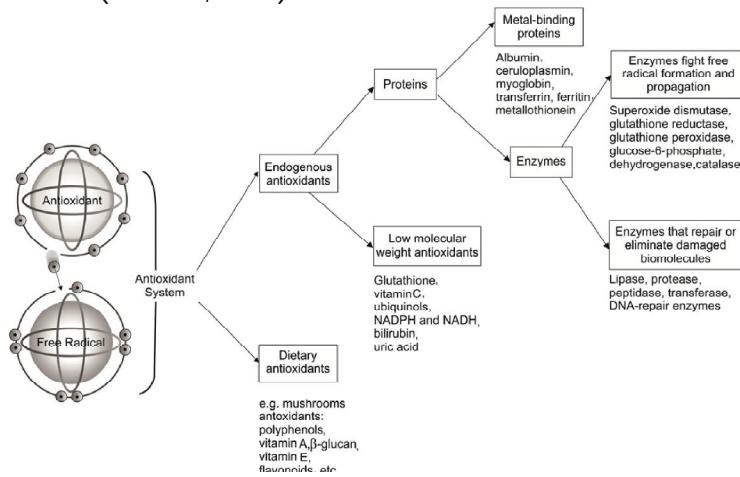
**Fig. 1. Endogenous and Exogenous Factors Inducing ROS Generation.**

The production of ROS can be reported by different external sources like tobacco, smoke, stress etc. or biproduct of electron transport of mitochondria or by oxidoreductase enzymes and metal catalyzed oxidation (Cederbaum et al., 2009). These reactive chemical compounds (free radicals) are components of cell especially cell wall or nucleic acid (DNA). They involve in important enzyme reaction and changing the chemical composition. By this action, the function of cell may lose or checked and results apoptosis or cellular senescence (Sanchez, 2017; Lakshmi et al., 2004; Cederbaum et al., 2009). So free radicles become a major agent of aging and degenerative diseases like immune system decline, cancer, liver disease, cardiac disease, diabetes, inflammation, kidney failure, stress etc. (Sanchez, 2017; Ma et al., 2018; Lakshmi et al., 2004; Alliwell, 1994) (Fig. 2).



**Fig. 2. Representation of human cell, which can be damaged by free radicals generated from internal and external sources. Neutralizing of free radicals by an antioxidant agent.**

Neutralizing agents of free radicals are specified as antioxidant agent which play necessary role in cell protection against ROS or Oxidative reaction through the exchange of their own electrons with free radicles. Antioxidant chemical compounds (Vitamin-A/Carotenoids, Vitamin-E/Tocopherol, Vitamin-C/Ascorbic acid, Polyphenol,  $\beta$ -Glucon, Glutathione etc.) provide most important intra-cellular defense against deleterious effects of ROS (Sanchez, 2017; Khatun et al., 2015). Antioxidant compounds may be two types- Endogenous and Exogenous (Fig. 3). Endogenous antioxidant agents are Proteins and some low molecular weight compounds like Uric acid, Bilirubin, Ubiquinols, Vitamin-C, Glutathione, NADPH, NADH etc. Exogenous antioxidant components are Dietary antioxidant which gain from external nutritional sources (Sanchez, 2017).



**Fig. 3. Representation of antioxidant system.**

During 1950s, "Free Radical Theory of Ageing" published by Harman and express the endogenous free oxygen radicals evolved to cause damages of cells. Free radical or reactive species such as ROS (Reactive Oxygen Species), RNS (Reactive Nitrogen Species), RCS (Reactive Carbon Species), RSS

(Reactive Sulfur Species) are influence the cell homeostasis and ageing. In all reactive species, ROS represent the most important and dominating category of living system and more than 90% production reported in eukaryotic cell by mitochondrial ETS (Electron Transport System) (Kozarski et al., 2015). In mushrooms (Edible & Non edible) species, several bioactive molecules are reported that shows high health promoting properties with different bioactivities like antiviral, antibacterial, antifungal, anticancerous, antiallergic, anti-inflammatory, immunostimulatory, cholesterol lowering, immunosuppressive and especially antioxidant. Antioxidant activities can also say life-saving performance of bioactive molecules. The capacity of antioxidant of mushroom is determined by mainly the amount of phenolic compounds in their extracts (Kosanic et al., 2013; Feleke and Doshi, 2017). In mushroom, main chemical is phenolic compounds like phenolic acid, tannin, lignin, stilbenes, flavonoids, hydroxybenzoic acid, hydroxycinnamic acid and polyphenols which perform a significant antioxidant activity in biological systems by inhibiting free radicals. Number of phenolic compounds in mushroom extracted as 1-6mg phenolics from 1g dried mushroom (Ma et al., 2018). Different type of methods evolved to measure the antioxidant efficiency. The working mechanism of these methods based on antioxidant defense system such as inhibition of free radicals which are able to induce cellular damages (Lakshmi et al., 2004). Two type of antioxidant availability have been seen i.e., synthetic antioxidant and natural antioxidant. Several natural antioxidants have been demonstrated by researchers from different natural products such as cereals, pulses, vegetables, fruits, leaves, roots, and Mushrooms. Natural antioxidant may consume by dietary and medicinal means (Lakshmi et al., 2004). Both type of antioxidant such as natural and synthetic are effective to reducing oxidative damage in human body by ROS. However, some synthetic antioxidants are susceptible to have toxic and carcinogenic effects like BHA (Butylated hydroxy anisole), BHT (Butylated hydroxy toluene), TBHQ (Tetra butylhydroquinone) EQ (Ethoxyquin) and PG (Propyl gallate). Therefore, the desired antioxidant is reported to natural antioxidant which has a natural origin. The development and utilization of natural antioxidant are more effective (Kosanic et al., 2012; Kozarski et al., 2015).

**Table 1. List of some mushroom species which has antioxidant activities.**

| S.N. | Mushroom Species               | Edibility | References   |
|------|--------------------------------|-----------|--|
| 1    | <i>Agaricus bisporus</i>       | Edible    | Ramkumar et al., 2010; Ma et al., 2018; He et al., 2012; Taofiq et al., 2016         |
| 2    | <i>Agaricus compestris</i>     | Edible    | Akata et al., 2019   |
| 3.   | <i>Agrocybeae grata</i>        | Edible    | Mujic et al., 2010   |
| 4.   | <i>Agrocybocylindracea</i>     | Edible    | Murcia et al., 2002  |
| 5.   | <i>Aleurodiscus vitellinus</i> | Edible    | Toledo et al., 2016  |
| 6.   | <i>Amanita crocea</i>          | Edible    | Alkan et al., 2020   |
| 7.   | <i>Amanita porphyria</i>       | Inedible  | Reis et al., 2011  |
| 8.   | <i>Amanita rubescens</i>       | Edible    | Kosanic et al., 2013   |
| 9.   | <i>Angelini sps</i>            | Edible    | Alkan et al., 2020   |
| 10.  | <i>Auricularia auricula</i>    | Edible    | He et al., 2012; Hussein et al., 2015; Boonsong et al., 2016; Obodai et al., 2014    |
| 11.  | <i>Boletus aestivalis</i>      | Edible    | Kosanic et al., 2012   |
| 12.  | <i>Boletus edulis</i>          | Edible    | Ma et al., 2018; Kosanic et al., 2012; Vamanu and Nita, 2013                         |
| 13.  | <i>Calocybe ambrosia</i>       | Edible    | Ma et al., 2018; Vishwakarma et al., 2016  |
| 14.  | <i>Calocybe indica</i>         | Edible    | Ramkumar et al., 2010; Ma et al., 2018; Vishwakarma et al., 2016                     |
| 15.  | <i>Cantharellus cibarius</i>   | Edible    | Kosanic et al., 2013; Ma et al., 2018; Ramesh and Pattar, 2010; Kosanic et al., 2013 |
| 16.  | <i>Cantharellus cinerius</i>   | Edible    | Kumari et al., 2011  |
| 17.  | <i>Cantharellus friesii</i>    | Edible    | Kumari et al., 2011  |

|     |                                   |          |  |
|-----|-----------------------------------|----------|--|
| 18. | <i>Cantharellus lutescens</i>     | Edible   | Murcia et al., 2002  |
| 19. | <i>Cantharellus subcibarius</i>   | Edible   | Kumari et al., 2011  |
| 20. | <i>Clavaria vermicularis</i>      | Edible   | Ramesh and Pattar, 2010  |
| 21. | <i>Collybia fusipes</i>           | Inedible | Reis et al., 2011  |
| 22. | <i>Coprinus comatus</i>           | Edible   | Akata et al., 2019   |
| 23. | <i>Cortinarius magellanicus</i>   | Edible   | Toledo et al., 2016  |
| 24. | <i>Craterellus cornucopioides</i> | Edible   | Ma et al., 2018  |
| 25. | <i>Cyclocybe cylindracea</i>      | Edible   | Alkan et al., 2020   |
| 26. | <i>Cytaria hariootii</i>          | Edible   | Toledo et al., 2016  |
| 27. | <i>Fistulina antarctica</i>       | Edible   | Toledo et al., 2016  |
| 28. | <i>Fistulina endoxantha</i>       | Inedible | Toledo et al., 2016  |
| 29. | <i>Flammulinavelutipes</i>        | Edible   | He et al., 2012  |
| 30. | <i>Fomitopsis spinicola</i>       | Inedible | Reis et al., 2011  |
| 31. | <i>Ganoderma lucidum</i>          | Edible   | Lakshmi et al., 2004; 55   |
| 32. | <i>Grifola frondosa</i>           | Edible   | Yeh et al., 2011   |
| 33. | <i>Grifola garga</i>              | Edible   | Toledo et al., 2016  |
| 34. | <i>Hebeloma sinapizans</i>        | Inedible | Reis et al., 2011  |
| 35. | <i>Hemileccinum dipilatum</i>     | Edible   | Alkan et al., 2020   |
| 36. | <i>Hericiumerinaceum</i>          | Edible   | Mujic et al., 2010   |
| 37. | <i>Hybsizus ulmarius</i>          | Edible   | Ramkumar et al., 2010  |
| 38. | <i>Hydnus repandum</i>            | Edible   | Murcia et al., 2002  |
| 39. | <i>Hydropus duseñii</i>           | Inedible | Toledo et al., 2016  |
| 40. | <i>Hygrocybe acuteconica</i>      | Inedible | Alkan et al., 2020   |
| 41. | <i>Hygrophorus marzuolus</i>      | Edible   | Ma et al., 2018  |
| 42. | <i>Infundibulicybe geotropa</i>   | Edible   | Sevindik et al., 2020  |
| 43. | <i>Inocybe splendens</i>          | Inedible | Reis et al., 2011  |
| 44. | <i>Lactarius deliciosus</i>       | Edible   | Ma et al., 2018; Alkan et al., 2020                                |
| 45. | <i>Lactarius hepaticus</i>        | Inedible | Reis et al., 2011  |
| 46. | <i>Lactarius piperatus</i>        | Inedible | Kosanic et al., 2013   |
| 47. | <i>Leccinum carpini</i>           | Edible   | Kosanic et al., 2012   |
| 48. | <i>Lentinus edodes</i>            | Edible   | Boonsong et al., 2016; He et al., 2012                             |
| 49. | <i>Lentinus sajor caju</i>        | Edible   | Hussein et al., 2015   |
| 50. | <i>Lentinus squarrosulus</i>      | Edible   | Hussein et al., 2015; Obodai et al., 2014                          |
| 51. | <i>Lentinus tigrinus</i>          | Inedible | Reis et al., 2011  |
| 52. | <i>Lepista edodes</i>             | Edible   | Murcia et al., 2002  |
| 53. | <i>Lepista nuda</i>               | Edible   | Murcia et al., 2002; Toledo et al., 2016                           |
| 54. | <i>Letinula edodes</i>            | Edible   | Ma et al., 2018; Taofiq et al., 2016; Mujic et al., 2010           |
| 55. | <i>Leucoagaricus leucothites</i>  | Edible   | Akata et al., 2019   |
| 56. | <i>Leucopaxillus giganteus</i>    | Edible   | Feleke and Doshi, 2017   |
| 57. | <i>Lycoperdon perlatum</i>        | Edible   | Ramesh and Pattar, 2010  |
| 58. | <i>Lycoperdon nutriforme</i>      | Edible   | Akata et al., 2019   |
| 59. | <i>Macrolepiota mastoides</i>     | Edible   | Akata et al., 2019   |
| 60. | <i>Macrolepiota procera</i>       | Edible   | Hussein et al., 2015; Akata et al., 2019; Vishwakarma et al., 2016 |
| 61. | <i>Marasmius oreades</i>          | Edible   | Ramesh and Pattar, 2010  |
| 62. | <i>Neoboletus erythropus</i>      | Edible   | Alkan et al., 2020   |
| 63. | <i>Panaeolus antillarum</i>       | Inedible | Dulay et al., 2015   |
| 64. | <i>Panus conchatus</i>            | Inedible | Hussein et al., 2015   |
| 65. | <i>Phellinus rimosus</i>          | Inedible | Ajith and Janardhanan, 2007  |

|      |                                  |           |  |
|------|----------------------------------|-----------|--|
| 66.  | <i>Piptoporusbetulinis</i>       | Inedible  | Reis et al., 2011  |
| 67.  | <i>Pleurotus citrinopileatus</i> | Edible    | Khatun et al., 2015  |
| 68.  | <i>Pleurotus djamor</i>          | Edible    | Ramkumar et al., 2010; Arbayah and Umi Kalsom, 2013  |
| 69.  | <i>Pleurotus eryngii</i>         | Edible    | Gasecka et al., 2016; Yildirim et al., 2012  |
| 70.  | <i>Pleurotus euos</i>            | Edible    | Ramkumar et al., 2010; Boonsong et al., 2016   |
| 71.  | <i>Pleurotus florida</i>         | Edible    | Ramkumar et al., 2010; Lakshmi et al., 2004; Khatun et al., 2015; Ajith and Janardhanan, 2007  |
| 72.  | <i>Pleurotus ostreatus</i>       | Medicinal | Taofiq et al., 2016; Obodai et al., 2014; Arbayah and Umi Kalsom, 2013; Ma et al., 2018; Chirinang and Intarapichet, 2009; Gasecka et al., 2016; Iwalokun et al., 2007 |
| 73.  | <i>Pleurotus platypus</i>        | Edible    | Ramkumar et al., 2010  |
| 74.  | <i>Pleurotus pulmonarius</i>     | Edible    | Arbayah and Umi Kalsom, 2013; Khatun et al., 2015; Ajith and Janardhanan, 2007; Nguyen et al., 2016; Ramesh and Pattar, 2010   |
| 75.  | <i>Pleurotus rimosus</i>         | Inedible  | Lakshmi et al., 2004   |
| 76.  | <i>Pleurotus sajor caju</i>      | Edible    | Boonsong et al., 2016; Ramkumar et al., 2010; Obodai et al., 2014; Lakshmi et al., 2004; Chirinang and Intarapichet, 2009  |
| 77.  | <i>Pleurotus tuber regium</i>    | Edible    | Obodai et al., 2014  |
| 78.  | <i>Pluteus murinus</i>           | Inedible  | Reis et al., 2011  |
| 79.  | <i>Polyporusdermoporus</i>       | Inedible  | Dore et al., 2014  |
| 80.  | <i>Polyporustenuiculus</i>       | Inedible  | Hussein et al., 2015   |
| 81.  | <i>Ramariaformosa</i>            | Edible    | Ramesh and Pattar, 2010  |
| 82.  | <i>Ramariapatagonica</i>         | Inedible  | Toledo et al., 2016  |
| 83.  | <i>Ramaria stricta</i>           | Edible    | Krupodorova and Sevindik, 2020   |
| 84.  | <i>Ramariasubalpina</i>          | Edible    | Acharya et al., 2017   |
| 85.  | <i>Russulaaurea</i>              | Edible    | Alkan et al., 2020   |
| 86.  | <i>Russulacyanoxantha</i>        | Edible    | Kosanic et al., 2013   |
| 87.  | <i>Russuladelica</i>             | Edible    | Turkoglu et al., 2017  |
| 88.  | <i>Russulaemetica</i>            | Inedible  | Reis et al., 2011  |
| 89.  | <i>Russulasanguinea</i>          | Edible    | Alkan et al., 2020   |
| 90.  | <i>Schizophyllum commune</i>     | Edible    | Arbayah and Umi Kalsom, 2013   |
| 91.  | <i>Suillusgranulatus</i>         | Edible    | Mushtaq et al., 2020   |
| 92.  | <i>Termitomycesaurantiacus</i>   | Edible    | Tibuhwa, 2012  |
| 93.  | <i>Termitomycesclypeatus</i>     | Edible    | Tibuhwa, 2012  |
| 94.  | <i>Termitomyceslestestui</i>     | Edible    | Tibuhwa, 2012  |
| 95.  | <i>Termitomycesmicrocarpus</i>   | Edible    | Tibuhwa, 2012  |
| 96.  | <i>Termitomyces robustus</i>     | Edible    | Obodai et al., 2014  |
| 97.  | <i>Termitomyces striatus</i>     | Edible    | Tibuhwa, 2012  |
| 98.  | <i>Termitomycestitanicus</i>     | Edible    | Tibuhwa, 2012  |
| 99.  | <i>Tuber aestivum</i>            | Edible    | Vishwakarma et al., 2016   |
| 100. | <i>Volvariellavolvacea</i>       | Edible    | Ramkumar et al., 2010; Boonsong et al., 2016   |

## CONCLUSION

Antioxidant is the chief and satisfactory component which increase the life of cell and delay the ageing process. Mushroom is the rich natural resource to fulfil the dietary balance of antioxidant. Plant are also rich in antioxidant properties but mushroom is better, easily available, suitable test with edibility, and economical in subject to human welfare. Phenolic compound is the chief antioxidant factor

of mushroom. In subject to knowledge of antioxidant properties of mushroom, much more research is required.

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